TECHNETIUM-99 IN WATER

1. SCOPE

1.1. This procedure describes a method to separate and measure technetium-99 in water.

1.2. This method does not address all aspects of safety, quality control, calibration or instrument set-up. However, enough detail is given for a trained radiochemist to achieve accurate and precise results for the analysis of the analyte(s) from the appropriate matrix, when incorporating the appropriate agency or laboratory safety, quality and laboratory control standards.

2. SUMMARY OF METHOD

2.1. Technetium-99 is separated from water samples using TEVA Resin. $^{99}$Tc is measured by liquid scintillation counting by either adding the resin directly to liquid scintillation cocktail or by stripping the technetium from the column and adding the $^{99}$Tc to the cocktail for counting. Each sample is analyzed with and without a $^{99}$Tc spike to determine chemical recovery. Alternately, the short-lived gamma emitter, $^{99m}$Tc, can be used as a tracer. The detection limit for this method is 1 pCi/L.

3. SIGNIFICANCE OF USE

3.1. This is a rapid, reliable method for measurement of $^{99}$Tc in environmental samples that is more cost-effective and efficient than traditional anion exchange, solvent extraction or precipitation techniques.

4. INTERFERENCES

4.1. Beta emitting radionuclides (including $^{14}$C, $^{32}$P, $^{35}$S, and $^{90}$Sr) and components that quench the liquid scintillation counting are effectively removed using Eichrom TEVA Resin. Tritium may follow the technetium due to the absorption of tritium-labeled compounds by the resin. Possible interference by tritium can be minimized by setting the $^{99}$Tc liquid scintillation counting window above the maximum energy for tritium beta particles.
4.2. Organic matter present in the sample can interfere by quenching during liquid scintillation counting. An Eichrom prefilter column is used to remove organics from the sample.

4.3. One liter of solution may be passed through a 2 mL prepacked TEVA Resin column without breakthrough of $^{99}$Tc.

4.4. Because $^{234}$Th has a beta decay with an energy in the $^{99}$Tc window, it is necessary to ensure complete decontamination from thorium. For samples high in $^{234}$Th it is recommended to follow procedure option #2, see 7.3.2. (Samples with high levels of natural uranium may contain significant $^{234}$Th.)

5. APPARATUS

- Column rack, Eichrom Part: AC-103
- Extension funnels, 25 mL, Eichrom Part: AC-120
- Glass beakers
- Liquid scintillation counter
- Liquid scintillation vials
- Watch glasses

6. REAGENTS

*Note:* Analytical grade or ACS grade reagents and trace metal grade (or equivalent) acids are recommended. *Evaluation of key reagents, such as aluminum nitrate and ammonium hydrogen phosphate, for contribution to method background levels from naturally occurring radioactive materials is recommended.*

| Deionized water, all reagents are prepared using deionized water
| Hydrofluoric acid (49%), concentrated HF -or- Sodium Fluoride, NaF
| Hydrogen peroxide (30%), concentrated H$_2$O$_2$
| Liquid Scintillation Cocktail
| Nitric acid (70%), concentrated HNO$_3$
| TEVA$^{®}$ resin, prepacked 2 mL columns, 100-150$\mu$m, Eichrom Part TE-C50-A
| Optional for removal of color from samples: Prefilter column -2mL prepacked columns, 100-150$\mu$m, Eichrom Part PF-C50-A

6.1. *Nitric acid (0.01M)* - Add 0.63mL of nitric acid to 950mL of water. Dilute to 1L with water.
6.2. *Nitric acid (0.02M)* - Add 1.25mL of nitric acid to 950mL of water. Dilute to 1L with water.

6.3. *Nitric acid (0.02M) / hydrofluoric acid solution (0.5M)* - Add 17.8mL of concentrated HF and 1.25mL of concentrated HNO₃ to 900mL water. Dilute to 1L with water.

6.4. *Nitric acid solution (0.1M)* - Add 6.3 mL of concentrated HNO₃ to 950mL of water. Dilute to 1 liter with water.

6.5. *Nitric acid solution (8M)* - Add 500 mL of concentrated HNO₃ to 250mL of water. Dilute to 1L with water.

6.6. *Nitric acid solution (1M)* - Add 62.5mL of concentrated HNO₃ to 900mL of water. Dilute to 1L with water.

7. **PROCEDURE**

7.1. Water Sample Preparation:

7.1.1. Measure the sample volume using a graduated cylinder (or equivalent) and transfer the volume to an appropriate size beaker.

7.1.2. Analyze each sample with and without adding ⁹⁹Tc spike to determine chemical recovery, or add appropriate yield tracer.

*Note: An alternative is to use ⁹⁹mTc as a tracer, measuring the short-lived gamma activity of ⁹⁹mTc (6.02 hour half-life) using gamma counting, allowing the ⁹⁹mTc to decay for approximately 1 week and then measure the Tc-99 beta using liquid scintillation counting.*

7.1.3. Add 10mL of 30wt% H₂O₂ (per liter of sample) into each beaker, stir and cover each beaker with a watch glass. Remove covers once the samples begin boiling.

7.1.4. Heat each beaker to about 90°C for 1 hour to oxidize Tc to Tc (VII), forming TcO₄⁻, oxidize some of the organics present, and destroy excess H₂O₂.

7.1.5. If bubbling due to decomposition of the hydrogen peroxide has not stopped as the sample cools, continue heating until bubbling has stopped. Stir occasionally with glass stirrer rod.

7.1.6. Allow beakers to cool to room temperature.

7.1.7. If the sample contains insoluble matter, filter the sample to remove solids prior to column loading.
7.2. TEVA Resin column preparation:

7.2.1. For each sample aliquot analyzed, place a TEVA Resin column in a column rack.

7.2.2. If organics that cause quenching are known to be present or may be present, place a prefilter column directly above each TEVA Resin column.

7.2.3. Place a beaker below each column, remove the bottom plug from each column and allow each column to drain.

7.2.4. Add 5mL of 0.1M HNO₃ into each column to condition the resin and allow to drain.

7.3. TEVA Resin column separation:

Note: The method describes the separation of Tc using 2mL prepacked TEVA resin columns. The method may also be performed using prepacked 2mL cartridges of prefilter (PF-R50-A) and TEVA resin (TE-R50-S) and the Eichrom vacuum box system (AR-BOX-24 or AR-BOX-12).

Note: If samples have high levels of $^{234}$Th (including samples high in natural uranium) then follow section 7.3.2 (Option #2).

7.3.1. Option #1

7.3.1.1. Transfer each water sample from step 7.1.7 into the appropriate Prefilter column (if required in step 7.2.2) and TEVA Resin column.

7.3.1.2. Allow the sample solution to drain through each set of columns.

7.3.1.3. Rinse the original beaker or container with the minimal volume of water required (depending on beaker size) and transfer this rinse to the appropriate column.

7.3.1.4. Allow the rinse solution to drain through each column.

7.3.1.5. Add 50mL of 0.01M HNO₃ directly into each column.

7.3.1.6. Allow the 0.01M HNO₃ rinse solution to drain through each column. Discard the rinses. **Proceed to section 7.4.**

7.3.2. Option #2 For samples containing high levels of Th-234
7.3.2.1. Transfer each water sample from step 7.1.7 into the appropriate Prefilter column (if required in step 7.2.2.) and TEVA Resin column.

7.3.2.2. Allow the sample solution to drain through each set of columns.

7.3.2.3. Rinse the original beaker or container with the minimal volume of water required (depending on beaker size) and transfer this rinse to the appropriate column.

7.3.2.4. Allow the rinse solution to drain through each column.

7.3.2.5. Add 5mL of 0.01M HNO₃ into each column. Discard the eluent.

7.3.2.6. Rinse column with 25mL of 0.5M HF/0.02M HNO₃.

*Note: Alternately, 40mL of 0.25M NaF/0.02M HNO₃ or 25 mL of 1M NaF/0.02M HNO₃ may be used. This step will remove any residual ²³⁴Th from the column.*

7.3.2.7. Rinse column with 5mL of 0.1M HNO₃ and discard the eluent. Proceed to section 7.4.

7.4. Counting preparation options:

7.4.1. Resin counting option (most rapid)

7.4.1.1. Remove the top frit and extrude the resin with a minimum volume of water into an LSC vial, or cut the ‘tip end’ of the column with a pair of scissors or blade and push the top frit with a glass rod so that the resin extrudes out into a LSC vial. Wash and transfer the rinse from each column with 3mL of 0.01M HNO₃ into the vial.

7.4.1.2. Add 10mL of the scintillation cocktail into each vial containing the resin. Cap the vial and shake well. GOTO 7.5.1.

*Note: Ultima Gold - XR™ or Ultima Gold - AB™ is suggested. Opti-Fluor™ or Insta-Gel XF™ or Ultima gold-LLT cocktails may also be used. Insta-Gel XF™ is less desirable from an environmental, waste disposal standpoint and is not required to fix the geometry since the extractant is stripped from the resin and homogeneously dispersed throughout the cocktail.*

7.4.2. Tc-99 column stripping option:
7.4.2.1. Place a clean labeled glass beaker or vial below each TEVA Resin column.

7.4.2.2. Add 20mL of 8M HNO₃ into each column to elute the Tc-99. Allow to drain. GOTO 7.4.2.3 or 7.4.2.4.

7.4.2.3. Direct Addition of Strip Solution to Cocktail option:

7.4.2.3.1. Add 1mL of the 8M HNO₃ from step 7.4.2.2 into a liquid scintillation vial.

7.4.2.3.2. Add 2mL of water, swirl to mix and add 10 mL of Ultima-Gold XR™ or Ultima-Gold AB™ scintillation cocktail.

Note: Ultima-Gold XR™ and Ultima-Gold AB™ are much more tolerant of acid than Opti-Fluor™, tolerating up to approximately 2 mL of 4M HNO₃ per 10 mL of cocktail.

7.4.2.3.3. Cap each vial, mix well and GOTO step 7.5.2.

7.4.2.4. Evaporation of strip solution option:

7.4.2.4.1. To minimize dilution, heat the beakers from step 7.4.2.2 gently, not greater than 80°C, until the volume in each beaker is about 10mL.

7.4.2.4.2. Transfer each solution from step 7.4.2.4.1 into a glass liquid scintillation vial.

7.4.2.4.3. Add two 2mL volumes of water to each beaker and transfer each rinse solution to the appropriate glass liquid scintillation vial.

7.4.2.4.4. Heat the vial gently (not greater than 80°C) until the solution volume is <1mL. Allow to cool, add 2mL of water and swirl to mix.

Note: Use a separate glass scintillation vial filled to exactly 0.5mL for volume comparison. If necessary, adjust the volume back up to 0.5mL with water. Do not go to dryness to avoid volatilizing Tc-99. If quenching problems occur, evaporate to a volume less than 0.5mL.

7.4.2.4.5. Add 10mL of liquid scintillation cocktail (or more volume, if desired) into each vial. Cap each vial, mix well and GOTO step 7.5.2.

7.5. Liquid scintillation counting:

7.5.1. Resin counting option:
7.5.1.1. Prepare a blank by preparing a vial containing the same amount of resin, water and cocktail as used in the resin counting method to determine background counts.

7.5.1.2. Prepare a $^{99}$Tc matrix standard by adding a known amount of $^{99}$Tc to a vial containing the same amount of resin, water and cocktail as used in the resin counting method to determine counting efficiency. GOTO step 7.5.3.

7.5.2. Tc-99 column stripping option:

7.5.2.1. Prepare a blank by preparing a vial containing the same amount of 8M HNO$_3$ and scintillation cocktail as used in the column stripping method to determine background counts.

7.5.2.2. Prepare a $^{99}$Tc matrix standard by adding a known amount of $^{99}$Tc to a vial containing the same amount of 8M HNO$_3$ and scintillation cocktail used in the $^{99}$Tc column stripping option to determine counting efficiency.

Note: If the evaporation option was used, evaporate the 8M HNO$_3$ used to prepare the blank and standard just as the samples were prepared.

7.5.3. Set up the scintillation counting window to measure from 20 Kev to 300 keV or alternate window optimized for the measurement of the $^{99}$Tc beta emission.

7.5.4. If the quenching between samples and standards is not similar, prepare a quench curve.

7.5.5. Count the vials the time required to obtain the counting statistics desired (typically 30 minutes to 1 hour) and to determine beta counts per minute.

7.5.6. Analyze a blank with each set of samples analyzed.

8. CALCULATIONS

Calculate the Tc-99 activity as follows:

$$\text{Sample dpm/L} = \frac{(S - B) \cdot D}{E \cdot V \cdot Y}$$

Where:

- $S$: Sample activity
- $B$: Blank activity
- $D$: Dilution factor
- $E$: Efficiency factor
- $V$: Volume of sample
- $Y$: Yield factor
S = sample counts/time in minutes, cpm
B = blank counts/time in minutes, cpm, steps 7.5.1.1 (resin counting option) or 7.5.2.1 (stripping option).
E = counting efficiency = measured cpm/dpm of Tc-99 matrix standard, steps 7.5.1.2 (resin option) or 7.5.2.2 (strip option)
V = sample volume, L
Y = yield = (spiked sample cpm - unspiked sample cpm) / E x Tc-99 spike activity, dpm

Note: If Tc-99m is used as a tracer, calculate the yield as follows:

\[
\text{Yield} = \frac{(C_s - B_s)}{E_s \times A_s}
\]

Where:

\(C_s\) = measured Tc-99m tracer, gamma cpm
\(B_s\) = background, gamma cpm
\(E_s\) = gamma counting efficiency for Tc-99m
\(A_s\) = Tc-99m tracer activity, dpm, corrected for decay from reference date

\(D = \) dilution factor = \(V_s / V_p\), included in calculation only if column strip method with direct addition to cocktail is used, step 7.4.2.3

Where:

\(V_s\) = strip volume, 20mL
\(V_p\) = volume of strip solution (12M HNO3) pipetted into cocktail, mL

Conversion of dpm/L to pCi/L: pCi/L = (dpm/L) / 2.22

9. PRECISION AND BIAS

9.1. Precision - A relative standard deviation of 3.4% at the 10,000 dpm level has been reported for procedure option #1 (7.3.1.)
9.2. Bias - A mean recovery of 92.5% has been reported for procedure option #1 (7.3.1.) Since results are corrected based on spike recovery, no significant bias exists for the method.

10. REFERENCES


1) Aliquot up to 1000mL of sample into glass beaker.
2) Add appropriate tracer and 10mL of 30% H₂O₂.
3) Place a watch glass on beaker and heat at 90°C for 1 hour.
4) Cool sample.
5) If necessary, filter sample to remove solids.
6) If necessary, use prefiter column to remove colored material.
7) Precondition TEVA column with 5mL 0.1M HNO₃.
8) Load sample on to TEVA. Allow liquid to drain to waste. Tc is retained.
9) Rinse sample beaker with 5mL water. Add beaker rinse to TEVA.
10) If sample contains Th-234/natural U, rinse TEVA with 25mL 0.5M HF-0.02M HNO₃.
11) Rinse TEVA with 50mL 0.01M HNO₃. Allow liquid to drain to waste.
12) Extrude resin into LSC vial or strip Tc with 20mL 8M HNO₃.